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(54) **POLYACRYLAMIDE PRECAST GELS FOR ELECTROPHORESIS, PROCESS FOR
PRODUCING THE SAME AND ELECTROPHORESIS METHOD BY USING THE GELS**

(57) A precast acrylamide electrophoresis gel is disclosed, which includes a gel buffer solution containing either Tris and glycine or Tris, glycine and any one or more than two of ampholytes, and a monobasic acid, titrated to a specified pH range. With the gels constituted as stated earlier, proteins can be successfully subjected to electrophoresis using the commonly available Laemmli's buffer system, while the DNA can be also satisfactorily run on the gel using the electrophoresis electrode solution containing Tris, chelating agent, and any one acid of acetic acid, phosphoric acid and boric acid.

In either of protein and DNA analyses, the gels even after stored under refrigeration more than six months are fully comparable in resolution and clearness of electrophoretic pattern to the gels just after being prepared. Where the DAN is subjected to electrophoresis using either the Laemmli's buffer or the Ornstein and Davis buffer, commonly used in electrophoresis of proteins, the gels even after storage under refrigeration over four months may exhibit the electrophoretic patterns comparable in resolution and clearness to the gels soon after being produced.

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Description

Technical Field

- 5 [0001] The present invention relates to a precast polyacrylamide gel adapted for use in electrophoresis separation, production method thereof and electrophoresis using the gel.

Background Art

- 10 [0002] The precast polyacrylamide gels for electrophoresis are extensively used as a basic investigative technique of detecting and quantitatively analyzing the chemical substances: proteins, nucleic acids, carbohydrates, and lipids necessary for building all living organisms in fields as diverse as biology, medicine, fisheries, veterinary medicine, and so on. Especially, a variety of precast polyacrylamide gels different in resolving power can be made easily by varying a gel recipe because the gels are substances synthesized artificially. Accordingly, it is possible to prepare in advance
15 the precast gels differing diversely in separation characteristic or resolution from one another. The artificial gels, since much saving the labor and effort for analytic procedure and excellent in uniformity and reproducibility, have helped well the productivity and quality control in the fields stated earlier. The artificial gels obtained on mass-production methods are required the increased shelf-life in preservation on their supply.

- [0003] Most of polyacrylamide gels for electrophoresis used extensively for analytic technique of proteins in the fields
20 of biochemistry and medicine are the system developed by Ornstein [L. Ornstein, Ann. N.Y. Acad. Sci. 121, 321-349 (1964)] and Davis [B. J. Davis, Ann. N.Y. Acad. Sci. 121, 404-427 (1964)] or that proposed by Laemmli [U.K. Laemmli, Nature, 227, 680 (1970)]. The gels have heretofore produced by the users or analysts in themselves or on themselves for private their use. In particular, the Laemmli system is most extensively used, since the molecular weights of proteins can be simply presumed by adding sodium dodecyl sulfate, contracted to "SDS" hereinafter, to gels or electrophoresis
25 electrode solutions. The Laemmli system consists of a gel buffer solution of tris(hydroxymethyl)aminomethane, contracted to "Tris" herein which is partially neutralized with hydrochloric acid, and an electrophoresis electrode solution, or Laemmli's electrophoresis electrode solution using Tris and glycine salt. The gel buffer solution in the system stated earlier, or Laemmli's gel buffer solution contains about 10-20mol% Tris partially neutralized to be adjusted to pH 8.8 with hydrochloric acid. At this pH of the Laemmli's gel buffer solution, however, amide groups are subjected to hydrolysis
30 with time that has elapsed. As the hydrolysis of gels proceeds even under low temperatures, the polyacrylamide gels come to contain partially anionic groups. As a result, the migration distances of proteins are made less while electrophoretic patterns become vague and therefore it is impossible to preserve the gels over a prolonged period.

- [0004] The following discussions will present the electrophoresis of nucleic acids. Recently remarkable development of molecular biology has come to need the analysis and preparation of deoxyribonucleic acid (DNA) and ribonucleic acid (RNA). The analysis of nucleic acids mostly depends on either the agarose electrophoresis separation process or the precast polyacrylamide gel electrophoresis separation process. As nucleic acids present strong negative charges in the neutral buffer solution while the mobility thereof depends on the molecular sieve effect of supportive matrix gels, either an about 0.3-2% agarose gel or an about 3.5-20% polyacrylamide gel is chiefly employed depending on the size of nucleic acids undergoing analysis.
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- [0005] The agarose gel, because of having a larger pore size, is often used for analytic separation of macromolecular nucleic acids. There are many types of agarose gels, which differ from one another in electroosmosis, gel strength, melting point, and so on. Moreover, because the agarose gels are naturally occurring materials, the differences in quality are frequently seen every a lot of products even the same gels produced in the same facility. For purifying DNA by virtue of the agarose gel electrophoresis separation process, any other purification procedures such as the phenol
40 extract, and so on must be considered because contamination with impurities in the agarose gels would inhibit enzyme activities of restriction enzymes, DNA polymerases, DNA synthetase, and so on.

- [0006] In contrast, the polyacrylamide gels with relatively small pore size are useful for isolation and analysis of nucleic acids of medium and/or low molecular weight. Because the polyacrylamide gels are artificially synthetic products, they are very high in chemical purity, and thus involving no major problems seen in the agarose gels. Moreover,
50 the polyacrylamide gels may be easily prepared in diverse types of gels, which are different in separation characteristics, depending on the desired gel recipe. Accordingly, it is possible to prepare in advance the precast gels differing diversely in separation characteristics from one another. The use of artificial gels mass-produced in advance in diverse separation characteristics, since much saving the labor and effort for analytic procedure and excellent in uniformity and reproducibility, will help well the productivity and quality control in the fields stated earlier. The artificial gels obtained on mass-
55 production methods are required the increased shelf-life in preservation on their supply. The most common buffer system for using agarose and polyacrylamide as the gel material in the electrophoresis separation of DNA is a continuous buffer system of around pH 7.8 to 8.3, in which a gel buffer solution is equal in composition with an electrophoresis electrode solution: a tris-acetic acid buffer, contracted to "TAE" herein, which contains either ethylenediaminetetraacetic

acid, contracted to "EDTA" or disodium ethylenediaminetetraacetate, contracted to "ETA"; a tris-boric acid buffer, abbreviated to "TBE"; and a tris-phosphoric acid, abbreviated to "TPE".

[0007] As stated earlier, the precast gels manufactured on mass-production methods must be able to maintain the long-lasting shelf-life in preservation throughout shipping and storage. Among the prior processes for manufacturing gels superior in shelf-life is a process for manufacturing a polyacrylamide gel having the increased shelf-life disclosed in Japanese Patent Laid-Open No. 184163/1992, which involves a gel buffer solution composed of Tris, ampholyte and acid to be storable for a prolonged period without loss of performance and also expanded remarkably in molecular weight range applicable for analytical electrophoresis. The polyacrylamide electrophoresis gel produced by the process recited just above is available for analysis using Laemmli's electrophoresis electrode solution. The polyacrylamide gel contains the ampholyte and has the pH of ranging from 4.0 to 7.5. In an example disclosed, the gel having a pH of ranging from 6.9 to 7.4 is produced, which is described to be stored under refrigeration for four months without variation in mobility of proteins.

[0008] The gel buffer solution is set high in neutralization rate of tris to depress the hydrolysis of the polyacrylamide gel, while added with the ampholyte to make gentle a variation in electric potential gradient neighboring a boundary between tris strong acid salt part and tris weak acid salt part, thereby coming to exhibit the wide range in the fractionated molecular weight of species undergoing measurement, which is comparable to the gels fabricated using the Laemmli's electrophoresis electrode solution.

[0009] Nevertheless, most polyacrylamide gels containing a gel buffer solution of the pH range 6.9 to 7.4 begin to vary in electrophoretic mobility of proteins after more than five months in storage under refrigeration has gone by. Moreover, the polyacrylamide gel in the form of slab gel is apt to change in shape of gel, tending to dislocate the glass plates. This results in major problems of rendering handling procedure on the electrophoretic analysis worse, causing the cracking of the gels due to the dislocation of the glass plates, and so on. Although it will be speculated that the gels may be improved in shelf life with the gel buffer solution of pH 6.8 or less, such gels have disadvantages of time-consuming for electrophoretic migration and also reducing in the resolution. Thus, none could not have succeed in developing a polyacrylamide gel that can exhibit the wide range in the fractionated molecular weight of species undergoing measurement, which is comparable to the gels fabricated using the Laemmli's gel buffer solution.

[0010] In addition, Published Japanese translations of PCT international publication No. 512907/1997 discloses a buffer system for a long-lasting precast electrophoresis gel wherein separation occurs at neutral pH. The gel buffer solution in the buffer system contains a mono organic amine or substituted amine with a pH near neutrality, titrated with hydrochloric acid to a pH of between about pH 6 and about pH 7. The electrophoresis electrode solution in the buffer system recited above contains a zwitterion selected from the group of MOPS, MES, ACES, MOPSO, TES, HEPES and TAPSO, titrated to a pH of about pH 7 with sodium hydroxide or organic base. Both of the gel buffer solution and the electrophoresis electrode solution are kept at neutral pH and in doing so the polyacrylamide gel itself is less subject to hydrolysis while proteins are separated with remaining completely reduced. The electrophoresis gel system recited earlier is described to have the increased useful shelf-life up to twelve months.

[0011] Nevertheless, the buffer system disclosed in Published Japanese translations of PCT international publication No. 512907/1997 requires using any special molecule markers, or molecules of known molecular weights. Moreover, the polyacrylamide in Published Japanese translations of PCT international publication No. 512907/1997 gets slower in the rate of electrophoretic migration when using any prior electrophoresis electrode solution, for example a commonly available Laemmli's electrophoresis electrode solution containing glycine. Thus, it is impossible to use the polyacrylamide for analytic separation of proteins.

[0012] The polyacrylamide controlled to a pH ranging from neutral to acid, as considerably resistant to hydrolysis, may serve well as the electrophoresis gel improved in shelf life. That is, the polyacrylamide set at a pH ranging from neutral to acid is prolonged in shelf life, and thus may be produced in advance by mass production methods to have diverse separation characteristics. If the polyacrylamide precast gel may be used combined with the electrophoresis electrode solution available for the electrophoresis of proteins to separate analytically DNA, or used combined with the electrophoresis electrode solution commonly available for the electrophoresis of nucleic acids to separate analytically DNA, it will be expected to realize the electrophoretic analysis with efficiency at low costs.

Disclosure of the Invention

[0013] A primary object of the present invention to solve the subject matter stated earlier is to provide a precast polyacrylamide gel that can be stored under refrigeration over more than six months with preserving the gel shape without loss of resolution of proteins, while coming to exhibit the wide range in the fractionated molecular weight of species undergoing measurement, which is comparable to the gels fabricated using the Laemmli's electrophoresis electrode solution.

[0014] A second object of the present invention to solve the subject matter stated earlier is to provide an electrophoresis method of separating and analyzing distinctly DNA by the use of a polyacrylamide electrophoresis gel storable

under refrigeration over more than six months without loss of performance, together with an electrophoresis electrode solution available commonly for the analytic separation of nucleic acids.

[0015] A third object of the present invention to solve the subject matter stated earlier is to provide an electrophoresis method of separating and analyzing distinctly DNA by the use of a polyacrylamide gel available for separation of proteins and storable under refrigeration over more than four months without loss of performance, together with an electrophoresis electrode solution for the analysis of proteins.

[0016] In an aspect of the present invention, a precast polyacrylamide gel adapted for use in gel electrophoresis is disclosed, in which an electrophoresis electrode solution is composed of an aqueous solution containing a tris(hydroxymethyl) aminomethane and an ampholyte, and wherein the tris(hydroxymethyl)aminomethane is present in a concentration of between 0.07mol/litre and 0.2mol/litre while the ampholyte is composed of glycine and at least one conjugate ampholyte, which has a basic dissociation constant of $8.3 < pK_b < 9.6$, the conjugate ampholyte added ranging in amount from 0.1 to 30mol% with respect to the glycine, concentrations of the conjugate ampholyte and the glycine ranging in all between 0.1 and 0.3mol/litre, and a pH range being between pH 6.0 and pH 6.8.

[0017] The precast polyacrylamide gel of the present invention can be stored under refrigeration for over six months without loss of resolution and gel configuration, and also exhibit the wide range in the fractionated molecular weight of species undergoing measurement, which is comparable to the gels fabricated using the Laemmli's electrophoresis electrode solution.

[0018] In another aspect of the present invention, there is disclosed that the conjugate ampholyte is an amino acid having the same number of anionic and cationic groups in a molecule.

[0019] In a further another aspect the present invention, there is disclosed that the conjugate ampholyte is any at least one selected from serine, glutamine, tryptophane, methionine and phenylalanine.

[0020] In another aspect of the present invention, a process for fabricating a precast polyacrylamide gel suitable for electrophoresis is disclosed, which is comprised of polymerizing an admixture of an acrylamide, polyfunctional crosslinking agent, water and a buffer solution of composition defined in the following (1) and (2) in the presence of a polymerization initiator under the condition of pH range 6.0 ~ 6.8:

- (1) a tris(hydroxymethyl)aminomethane is present in a concentration of between 0.07mol/litre and 0.2mol/litre; and
- (2) an ampholyte is composed of glycine and at least one conjugate ampholyte,

- (a) the conjugate ampholyte has a basic dissociation constant of $8.3 < pK_b < 9.6$,
- (b) the conjugate ampholyte added ranges in amount from 0.1 to 30mol% with respect to the glycine, and
- (c) a total concentration of the conjugate ampholyte and the glycine ranges between 0.1 and 0.3mol/litre.

[0021] According to the process for fabricating a precast polyacrylamide electrophoresis gel as stated earlier, the improved precast polyacrylamide electrophoresis gel may be obtained with efficiency, which is increased in the useful shelf life where the gel can be stored for over six months without loss of resolution and gel configuration.

[0022] In another aspect of the present invention, an use of the precast polyacrylamide electrophoresis gel is disclosed, in which the analytic separation of proteins is performed using the electrophoresis electrode solution containing glycine and tris(hydroxymethyl) aminomethane added with dodecyl sulfate.

[0023] The use of the precast polyacrylamide electrophoresis gel as stated just above makes it possible to realize the wide range in the fractionated molecular weight of species undergoing measurement, which is comparable to the gels fabricated using the Laemmli's gel electrophoresis electrode solution.

[0024] In a further another aspect of the present invention, an electrophoresis system for DNA is disclosed, which makes use of an electrophoresis electrode solution containing tris(hydroxymethyl) aminomethane and a precast polyacrylamide gel characterized by the following (1) and (2);

- (1) a gel buffer solution contained in the precast polyacrylamide gel contains the tris(hydroxymethyl)aminomethane, at least one ampholyte rendering glycine indispensable, and a monobasic acid, titrated to a pH of between pH 6.0 and pH 7.5, and
- (2) the ampholyte contained in the gel buffer solution has a basic dissociation constant of $8.3 < pK_b < 9.6$, and includes an amino acid having the same number of anionic and cationic groups in a molecule.

[0025] In another aspect of the present invention, there is disclosed that the gel buffer solution contains the monobasic acid consisting of at least any one of hydrochloric acid and acetic acid, titrated to a pH of between pH 6.0 and pH 6.5 to increase the useful shelf life of a precast gel.

[0026] In another aspect of the present invention, there is disclosed that the gel buffer solution contains the monobasic acid consisting of at least any one of hydrochloric acid and acetic acid, titrated to a pH of between pH 6.5 and pH 7.5 to increase the useful shelf life of a precast gel.

[0027] In another aspect of the present invention, an electrophoresis electrode solution is disclosed which contains, aside from tris(hydroxymethyl)aminomethane, components defined in the following (1) and (2), titrated to a pH in a pH range 7.8~8.3:

- (1) any one of acetic acid, phosphoric acid and boric acid; and
- (2) a chelating agent of disodium ethylenediaminetetraacetate.

[0028] In a further another aspect of the present invention, there is disclosed the electrophoresis electrode solution containing the monobasic acid of boric acid.

[0029] In another aspect of the present invention, there is disclosed the electrophoresis electrode solution containing an ampholyte, aside from tris(hydroxymethyl)aminomethane, and titrated to a pH in a pH range 7.8 ~8.3.

[0030] In another aspect of the present invention, there is disclosed the electrophoresis electrode solution containing the ampholyte of glycine.

[0031] In a further another aspect of the present invention, there is disclosed the electrophoresis electrode solution containing dodecyl sulfate, in addition to the ampholyte of glycine.

Best Mode for Carrying Out the Invention

[0032] In accordance with the present invention, a gel adapted for use in analytic separation of proteins is provided, which is enhanced in the useful shelf-life thereof. The gel contains therein a buffer solution composed of Tris, glycine and at least one conjugate ampholyte, the Tris ranging in concentration in the gel from 0.07mol/litre to 0.2mol/litre, preferably from 0.08mol/litre to 0.1mol/litre. The limited concentration range recited above results in inclusion of leading ions enough in amount to ensure the electrophoresis time comparable to the gels fabricated using the Laemmli's gel buffer solution. With the Tris less in concentration in the gel than 0.07mol/litre, the electrophoretic pattern becomes overall too vague to avail for the analytic separation of proteins. In contrast, even if the concentration of the Tris in the gel is over 0.2mol/litre, the concentration of the leading ion in the gel becomes too high, thereby prolonging the electrophoresis time, making the operational efficiency of the analysis worse.

[0033] The ampholyte is composed of glycine and at least one conjugate ampholyte that has the basic dissociation constant of $8.3 < pK_b < 9.6$, preferably $9.0 < pK_b < 9.6$. Only glycine by itself, especially when the gel buffer solution is at pH 7 or less, can not realize the wide range in the fractionated molecular weight of species undergoing measurement, which is comparable to the gels fabricated using the Laemmli's gel buffer solution. On the other hand, with the conjugate ampholyte of $pK_b < 8.3$, the ampholyte is lower in pK_b than the Tris, and for the sake of which the gel buffer solution is varied in pH to make the fractionated molecular weight range much broaden. In contrast, if the conjugate ampholyte is in the condition of $pK_b > 9.6$, the ampholyte exceeds the glycine in basic dissociation constant, making no contribution to helping ensure the gentle potential gradient. To cope with this, when the conjugate ampholyte is selected from the group of serine, tryptophane, phenylalanine, and so on satisfying $9.0 < pK_b < 9.6$, the ampholytes migrate consecutively towards the anode side to their respective positions on the basis of the pK_b value, with the smallest pK_b migrating to the position most neighboring the anode, so that the gentle potential gradient may be established in the gel. Thus, the precast polyacrylamide gel of the present invention succeeds in realizing the wide range in the fractionated molecular weight of species undergoing measurement, which is comparable to the gels fabricated using the Laemmli's gel buffer solution.

[0034] According to the present invention, the amount of the conjugate ampholytes added is from 0.1 to 30mol%, preferably from 1 to 20mol%, with respect to glycine. When the conjugate ampholyte added is less in amount than 0.1mol%, the precast polyacrylamide gel fails in realizing the wide range in the fractionated molecular weight of species undergoing measurement, which is comparable to the gels fabricated using the Laemmli's gel buffer solution. To the contrary, if the conjugate ampholyte is added over 30mol%, the electrophoretic image appearing on the gel becomes too vague to have the practical use.

[0035] The total concentration of the conjugate ampholyte and glycine is in the range of from 0.1mol/litre to 0.3mol/litre, preferably from 0.1 to 0.2mol/litre. Even when the ampholyte is either less than 0.1mol/litre or more than 0.3mol/litre, the electrophoretic pattern on the gel also becomes vague to be unavailable for practical use thereof.

[0036] The ampholytes used in the present invention preferably has the same number of anionic and cationic groups in a molecule. All the anionic and-cationic groups dissociate into ions in the gel buffer solution so that the ampholytes becomes electrical neutrality. At a pH higher than pK_b , only the anionic groups dissociate while the cationic groups are less subject to the dissociation so that the ampholytes is charged negatively. Thus, it will be understood that the ampholytes behave substantially like weak monobasic acid.

[0037] The given pH of the gel buffer solution for the precast polyacrylamide gel of the present invention, the use and production thereof is adjusted to from 6.0 to 6.8, preferably at 6.3. When the buffer solution is above pH 6.8, the polyacrylamide is susceptible to hydrolysis, and thus can not be stored for a long period without loss of performance.

In contrast, if the gel buffer solution is less than 6.0 in pH, the polyacrylamide gel may keep the increased shelf-life while the electrophoretic patterns of proteins are liable to become too vague to have the practical use. The polyacrylamide gel with the gel buffer solution adjusted to pH 6.8 can be stored under refrigeration over six months with no loss of performance. When controlled to just pH 6.3, the gels obtained are remarkably retarded in the rate of hydrolysis compared with the pH 6.8, because the pH 6.3 is near to the pH of the acrylamide itself. This makes it possible to store the gels for over a year with no change or deterioration in gel shape and electrophoretic pattern due to acrylamide hydrolysis.

[0038] The use of the precast polyacrylamide gels according to the present invention is an analytic separation of proteins, which employs the electrophoresis electrode solution containing Tris and glycine, preferably containing Tris of 0.025mol/litre, glycine of 0.192mol/litre and SDS of 0.1% by weight. The electrophoresis electrode solution, because of having the same composition found in the extensively used Laemmli's gel buffer solution, serves well for the analytic separation of proteins with inexpensive cost and efficiency. Selection of an ampholyte, for example MOPS rather than glycine for the electrophoresis electrode solution may also serve for the analytic separation of proteins. Nevertheless, the fractionated molecular weight range performed by the gel comes into broadening, compared with the gel fabricated using the Laemmli's gel buffer solution used extensively.

[0039] The precast polyacrylamide gel of the present invention may be produced in the same fashion as the production of the prior polyacrylamide electrophoresis gels, excepting pH regulation and addition of ampholytes. For instance, N,N'-methylene bis-acrylamide, contracted to "BIS" herein, is among the most extensively used water-soluble divinyl compounds, which are able to conjugate with the acrylamide for cross-linking the acrylamide. Any other suitable cross-linker may be utilized such as N,N'-diallyltartardiamide, which is a kind of the commonly available water-soluble divinyl compounds.

[0040] An aqueous monomer solution used in the precast polyacrylamide gel of the present invention may include a water-soluble polymer such as agarose, polyacrylamide, polyvinyl alcohol, polyvinyl pyrrolidone, polyethylene oxide, polymethyl vinyl ether, and so on to endow the gel with elasticity and increase the gel strength. Alternatively, the monomer solution may be copolymerized with other monomers.

[0041] Copolymerization of monomers in the production of the precast polyacrylamide gel of the present invention takes place with radicals resulting from either polymerization initiator or exposure to any of ultraviolet light and ionizing radiation. The polymerization initiator is preferably, but limited to a polymerization initiator of redox system in which a peroxide such as ammonium persulfate, and so on is simultaneously used with a reducing agent such as N,N,N',N'-tetramethylethylenediamine, contracted to TEMED herein, and so on. The peroxide and reducing agent are used in the range of from 0.05 to 5% by weight/volume based on the total monomer. Copolymerization temperature, although unspecified as long as the initiator is allowed to serve well, is preferably in the range of from 15 to 50°C.

[0042] The following description will present the electrophoresis separation for DNA in which the precast polyacrylamide gel constructed according to the present invention to have the increased shelf-life is used in combination with the electrophoresis electrode buffer solution commonly used for the analytic separation of nucleic acid.

[0043] The gel buffer solution contains Tris, any one or more than two of ampholytes including glycine, and a monobasic acid. Preferably, the gel buffer solution contains Tris, glycine, an amino acid having the same number of anionic and cationic groups in a molecule of $8.3 < \text{pKb} < 9.6$ in basic dissociation constant, hydrochloric acid and acetic acid. The gel buffer solution without any one or more than two of ampholytes including glycine, when made acidic in pH, gets Tris to rise in neutralization rate, causing much more variation in the potential gradient at an area neighboring a boundary between the tris strong acid salt part and the tris weak acid salt part. This is apt to incur focusing of separated molecules nearby the boundary area, rendering the fractionated molecular weight range much reduced. Besides, the electrophoresis time is prolonged so that the separation operation is intolerably retarded in efficiency.

[0044] Since glycine may serve well to make gentle the change in potential gradient at the area near the boundary between the tris strong acid salt part and the tris weak acid salt part in the electrophoresis gel, it is inevitable for realizing the wide range in the fractionated molecular weight of species undergoing measurement, which is comparable to the gels fabricated using either the TBE or the Ornstein and Davis gel buffer solution. Nevertheless, even if the gel buffer solution is made much acid in pH, glycine can not control the change in potential gradient alone. To cope with this, it is desired in the gel buffer solution to conjugate glycine with any amino acid having the same number of anionic and cationic groups in a molecule of $8.3 < \text{pKb} < 9.6$ in basic dissociation constant. This makes it possible to render the potential gradient in the electrophoresis gel gentle.

[0045] The amino acids conjugated with glycine have the basic dissociation constant pKb of $8.3 < \text{pKb} < 9.6$, more preferably of $9.0 < \text{pKb} < 9.6$. In case where the amino acids conjugated with glycine has $\text{pKb} < 8.3$, the amino acids are lower in pKb than the Tris, and for the sake of which the gel buffer solution is varied in pH to make the fractionated molecular weight range much broadening. Conversely, when glycine is conjugated with the amino acids of $\text{pKb} > 9.6$, the amino acids exceed the glycine in pKb , making no contribution to helping ensure the gentle potential gradient. Where the amino acids of $9.0 < \text{pKb} < 9.6$ are used in the gel buffer solution, they migrate consecutively towards the anode side to their respective positions on the basis of the pKb value, with the smallest pKb migrating to the position

most neighboring the anode, so that the gentle potential gradient may be established in the gel. Among the amino acids satisfying $9.0 < pK_b < 9.6$ are serine, tryptophane, phenylalanine, and so on. According to the present invention, serine is preferred.

[0046] Moreover, it is desirable that the amino acids conjugated with glycine have the same number of anionic and cationic groups in a molecule. At a pH somewhat lower than pK_b , all the anionic and cationic groups dissociate completely into ions so that the amino acids takes on electric neutrality. In contrast, at a pH higher than pK_b , only the anionic groups dissociate whereas the cationic groups are retarded in dissociation, so that the amino acids are charged negatively. Thus, the amino acids exhibit substantially the behavior as monobasic weak acid to be allowed to use them as quasi-weak acid. Previously adding any quasi-weak acid to the gel buffer solution results in lowering an electric resistance of the trisweak acid salt part in the electrophoresis gel. Under the presence of strong acid radicals, in addition, the pH is so low that the amino acids become electrically neutral, thus making no contribution to electric conduction.

[0047] The pH of the gel buffer solution is in the range of from 6.0 to 7.5, preferably from 6.0 to 6.5. With the pH above 7.5, the polyacrylamide gels, because of much subject to hydrolysis with time that has lapsed, are limited in shelf-life thereof. Conversely, the pH below 6.0, although increasing the useful shelf-life of the polyacrylamide gels, raises the neutralization rate of Tris to the extent that might impair the effects of the buffer system. In order to increase the useful shelf-life of the precast polyacrylamide gels, it is preferable to get the pH of the gel buffer solution to near the pH of the acrylamide per se.

[0048] On the other hand, the electrophoresis electrode solution contains Tris, chelating agent, and any one acid of acetic acid, phosphoric acid and boric acid, preferably boric acid. With the gel buffer solution adjusted to a pH in the pH range of from 6.0 to 6.5, the Ornstein and Davis electrophoresis electrode solution is not practical for the analytic separation of DNA because the electrophoretic patterns obtained become vague. The TBA employed in the present invention contains Tris of 0.0892mol/litre, boric acid of 0.0890mol/litre and ETA of 0.0025mol/litre, preferably titrated to pH 8.3. The buffer system of the composition stated just earlier is the TBE extensively used for analytic separation of nucleic acid, and therefore may serve well for the analysis with inexpensive cost and efficiency. Moreover, the buffer system stated earlier makes it possible to finish the electrophoresis within a limited short period of time while ensuring the very clear patterns. Either of TAE and TPE, although enabling the analytic separation of DNA, needs plenty of time to conduct the electrophoresis while becoming worse in the analytic efficiency, compared with the TBE.

[0049] The following description will present the electrophoresis separation for DNA in which the precast polyacrylamide gel constructed according to the present invention to increase in the useful shelf-life is used in combination with the electrophoresis electrode buffer solution commonly used for the electrophoresis of proteins.

[0050] The gel buffer solution contains Tris, any one or more than two of ampholytes including glycine, and a monobasic acid. Preferably, the gel buffer solution contains Tris, glycine, an amino acid having the same number of anionic and cationic groups in a molecule of $8.3 < pK_b < 9.6$ in basic dissociation constant, hydrochloric acid and acetic acid. The gel buffer solution without any one or more than two of ampholytes including glycine, when made acidic in pH, gets Tris to rise in neutralization rate, causing much more variation in the potential gradient at an area neighboring a boundary between the tris strong acid salt part and the tris weak acid salt part. This is apt to incur focusing of separated molecules undergoing analysis nearby the boundary area, rendering the fractionated molecular weight range much reduced. Besides, the electrophoresis time is prolonged so that the separation operation is intolerably retarded in efficiency.

[0051] Since glycine may serve well to make gentle the change in potential gradient at the area near the boundary between the tris strong acid salt part and the tris weak acid salt part even in the low pH gel buffer solution, the existence of glycine in the gel buffer solution is inevitable for realizing the wide range in the fractionated molecular weight of species undergoing measurement, which is comparable to the gels fabricated using either the TBE or the Ornstein and Davis gel buffer solution. Nevertheless, when the gel buffer solution is made much acid in pH, glycine can not control the change in potential gradient alone. To cope with this, it is desired in the gel buffer solution to conjugate glycine with any amino acid having the same number of anionic and cationic groups in a molecule of $8.3 < pK_b < 9.6$ in basic dissociation constant. This makes it possible to render the potential gradient in the electrophoresis gel gentle.

[0052] The amino acids to be conjugated with glycine have the basic dissociation constant pK_b of $8.3 < pK_b < 9.6$, more preferably of $9.0 < pK_b < 9.6$. In case where the amino acids conjugated with glycine has $pK_b < 8.3$, the amino acids are lower in pK_b than the Tris, and for the sake of which the gel buffer solution is varied in pH to make the fractionated molecular weight range much broadening. Conversely, when glycine is conjugated with the amino acids of $pK_b > 9.6$, the amino acids exceed the glycine in pK_b , making no contribution to helping ensure the gentle potential gradient. Where the amino acids of $9.0 < pK_b < 9.6$ are used in the gel buffer solution, they migrate consecutively towards the anode side to their respective positions on the basis of the pK_b value, with the smallest pK_b migrating to the position most neighboring the anode, so that the gentle potential gradient may be established in the gel. Among the amino acids satisfying $9.0 < pK_b < 9.6$ are serine, tryptophane, phenylalanine, and so on. According to the present invention, serine is preferred.

[0053] Moreover, it is desirable that the amino acids conjugated with glycine have the same number of anionic and

cationic groups in a molecule. At the pH somewhat lower than pK_b , all the anionic and cationic groups dissociate completely into ions so that the amino acids takes on electric neutrality. In contrast, at the pH higher than pK_b , only the anionic groups dissociate whereas the cationic groups are retarded in dissociation, so that the amino acids are charged negatively. Thus, the amino acids exhibit substantially the behavior as monobasic weak acid to be allowed to use them as quasi-weak acid. Previously adding any quasi-weak acid to the gel buffer solution results in lowering an electric resistance of the trisweak acid salt part in the electrophoresis gel. Under the presence of strong acid radicals, in addition, the pH is so low that the amino acids become electrically neutral, thus making no contribution to electric conduction.

[0054] The pH of the gel buffer solution is in the range of from 6.5 to 7.5, preferably from 6.5 to 7.0. With the pH above 7.5, the polyacrylamide gels, because of much subject to hydrolysis with time that has lapsed, are limited in shelf-life thereof. Conversely, the pH below 6.5, although increasing the useful shelf-life of the polyacrylamide gels, results in unsatisfactory electrophoresis patterns.

[0055] The electrophoresis electrode solution employable in the present invention is the Omstein and Davis electrophoresis electrode solution containing 0.025mol/litre Tris and 0.192mol/litre glycine, titrated to the pH 8.3. The buffer solution composed as stated earlier is the electrophoresis electrode solution extensively used in the analytic separation of proteins and, therefore, may serve well with good efficiency to obtain very clear electrophoresis results.

[0056] For the electrophoresis electrode solution of the present invention, alternatively, the Laemmli's electrophoresis electrode solution commonly used in the protein analyses may be also adopted, which contains 0.025mol/litre Tris, 0.192mol/litre glycine and 0.1%(w/v) SDS. Thus, both the analytic separations of DNA and proteins may be run on the same electrophoresis electrode solution. This contributes to the efficient operation and cost reduction.

[0057] While the present invention will be explained with reference to the following examples, it should be understood that such examples are not restrictive, but illustrative purposes only.

-Example 1-

[0058] Arranged between a rectangular glass sheet of 12cm in width and 10cm long and another glass sheet of the same dimension made at an upper end thereof with a recess were a spacer of 1mm in thickness and a silicon seal to keep monomer solution against leakage. They were assembled into a glass plate cell. Monomer solutions were prepared, each of which was composed of a 10% acrylamide (%T: "total monomer concentration in the solution"), a 5% BIS (%C: "concentration of BIS"), and any one of buffer solutions composed as shown in Table 1 below. The monomer solutions were added and blended with 0.0025mol/litre APS and 0.0075mol/litre TEMED. The resultant mixture was poured into the glass plate cell where polymerization was allowed at 25 °C to yield a polyacrylamide gel for electrophoresis.

[0059] The protein marker, or commercially available protein of known molecular weight, was electrophoresed on the polyacrylamide electrophoresis gel obtained as stated earlier to measure the migration distance. The protein marker was first treated with 2-mercaptoethanol and SDS and further added with a 0.5wt% bromphenol blue, contracted to BPB herein, and 7wt% glycerol to be used for a test. The electrophoresis electrode solution had the composition of Laemmli system composed of 0.025mol/litre Tris, 0.192mol/litre glycine, 0.1wt% SDS. The electrophoresis was done using a constant current of 20mA. Electric conduction was halted just when the dye front of BPB had reached 5mm above the bottom of the gel. The staining was carried out using 0.25vol% Coomassie Brilliant Blue, contracted to CBB hereinafter, G-250, 10vol% acetic acid, and 30vol% methanol solution with shaking for 45minutes while the destaining was performed using 7vol% acetic acid, 3vol% methanol solution with shaking for 90minutes. Table 2 shows the mobility just after being produced while Table 3 is the mobility after stored at 5 °C for six months. The mobility is defined as the migration distance of protein in percent:

$$(\text{Mobility } \%) = (\text{Distance of each band from the lower end of well}) / (\text{Distance of the dye front of BPB from the lower end of well}) \times 100$$

Table 1

[Compositions of gel buffer solutions]					
	Sample -1	Sample -2	Comp. -1	Comp. -2	Comp. -3
Tris (mol/litre)	0.082	0.080	0.375	0.075	0.082
Glycine (mol/litre)	0.167	0.162	--	0.192	0.182

Table 1 (continued)

[Compositions of gel buffer solutions]					
	Sample -1	Sample -2	Comp. -1	Comp. -2	Comp. -3
Serine (mol/litre)	0.025	0.03	--	--	--
Tricine (mol/litre)	--	--	--	--	0.01
pH	6.8	6.3	8.8	6.8	6.8

Table 2

[Mobility(%) just after being produced]					
	Sample -1	Sample -2	Comp. -1	Comp. -2	Comp. -3
Protein Mol. wt. 94,000	23.81	22.49	23.07	26.78	24.49
Protein Mol. wt. 43,000	50.79	50.24	50.48	53.55	48.47
Protein Mol. wt. 20,100	91.53	94.26	92.91	--	77.04
Phoretic result	Clear	Clear	Clear	Clear	Clear

Table 3

[Mobility(%) after being stored at 5 °C for six months]					
	Sample -1	Sample -2	Comp. -1	Comp. -2	Comp. -3
Protein Mol. wt. 94,000	26.67	28.43	17.10	25.51	24.02
Protein Mol. wt. 43,000	52.31	55.39	42.20	54.08	47.98
Protein Mol. wt. 20,100	94.36	96.08	88.99	--	76.40
Phoretic result	Clear	Clear	Vague	Clear	Clear

[0060] The gel fabricated using the Laemmli's gel buffer solution less in neutralization rate caused by acids in the gel buffer solution was reduced in mobility after stored for a long period because of a high pH, became worse in resolution. With the gel buffer solution in which Tris is raised in neutralization rate while the ampholyte is composed of only glycine, although few reduction in mobility could discerned even after long storage, no small molecule range was found. Moreover, the gel buffer solution adjusted to the pH 6.8 and added with tricine as a conjugate ampholyte of 8.3 > pKb together with glycine made the fractionated molecular weight range much broadening.

[0061] In contrast, when the conjugate ampholyte of $9.0 < \text{pKb} < 9.6$, or serine was employed together with glycine, the ampholytes migrated consecutively towards the anode side to their respective positions on the basis of the pKb value, with the smallest pKb migrating to the position most neighboring the anode, so that the gentle potential gradient was established in the gel. Thus, even when Tris was raised in neutralization rate while the buffer solution was titrated to either pH 6.8 or pH 6.3, the gel succeeded in realizing the wide range in the fractionated molecular weight of species undergoing measurement, which is comparable to the gels fabricated using the Laemmli's gel buffer solution. Besides, no reduction in mobility was seen even after long storage.

-Example 2-

[0062] Ten-percent polyacrylamide gels were produced using the same glass plate cell as in Example 1. The monomer solutions used were prepared, each of which was composed of a 10% acrylamide (%T), a 5% BIS (%C), and any one of buffer solutions composed as shown in Table 4 below. The polyacrylamide gels obtained were stored at 5°C for twelve months. The glass plate cells after being stored were subjected to vertical tensile force by the use of a Tensilon Universal testing machine (Orientec Type U-2129) to measure tensile strength, the results of which are shown in Table 5.

Table 4

[Compositions of gel buffer solutions]			
	Sample-3	Sample-4	Comp.-4
Tris (mol/litre)	0.082	0.080	0.09375
Glycine (mol/litre)	0.167	0.162	0.192
Serine (mol/litre)	0.025	0.03	--
Tricine (mol/litre)	--	--	--
pH	6.3	6.8	7.5

Table 5 [Tensile strength(kgf) of the glass plate cell]

	Soon after produced	After 2 months at 5°C	After 4 months at 5°C	After 6 months at 5°C
Sample-3	1.85	1.83	1.88	1.83
Sample-4	1.83	1.80	1.78	1.73
Comp.-4	1.80	0.65	0.52	--

	After 8 months at 5°C	After 10 months at 5°C	After 12 months at 5°C
Sample-3	1.85	1.85	1.83
Sample-4	0.85	0.388	--
Comp.-4	--	--	--

[0063] Lowering pH in the gel buffer solution contributed to keeping not only the mobility of proteins but also the shape of the precast gel stable. As well known, the polyacrylamide gel is in general apt to swell readily due to hydrolysis, so that the precast gel becomes much subject to crack in the glass plate cell. At the pH 7.5, since the gel was liable to be much subjected to hydrolysis, the tensile strength causing the crack in the glass plate cell fell to about one third after stored at 5 °C for two months. Conversely, at the pH 6.8, the gel in the glass plate cell continued keeping as high tensile strength as just after produced, even after stored at 5°C up to six months. When controlled to the pH 6.3, the gels obtained were remarkably retarded in the rate of hydrolysis compared with the pH 6.8, because the pH 6.3 was near to the pH of the acrylamide itself. This made it possible to store the gels for over a year without no change or deterioration in gel shape.

-Example 3-

[0064] A rectangular glass sheet of 12cm in width and 10cm long and another glass sheet of the same dimension made at an upper end thereof with a recess were assembled with a spacer of 1mm in thickness interposed between the confronting glass sheets, providing a glass plate cell. Monomer solutions were prepared, each of which was composed of a 10% acrylamide (%T), a 5% BIS (%C), and any one of buffer solutions composed as shown in Table 6 below. The monomer solutions were added and blended with 400ppm of APS and 400ppm of tetramethylethylenedi-

amine, contracted to TEMED. The resultant mixture was poured into the glass plate cell where polymerization was allowed in a usual way to yield a polyacrylamide gel for electrophoresis.

[0065] A commercially available marker DNA of known molecular weights was subjected to electrophoresis using the polyacrylamide gel obtained as stated earlier. The electrophoresis results were satisfactory in migration distance and clearness of bands. The marker DNA, prior to use for electrophoresis test, was diluted one hundred times with a buffer solution composed of a 10m mol/litre Tris including 30%(w/v) sucrose and BPB of stainable level, 1m mol/litre ETA and 20m mol/litre sodium chloride solution, titrated to the pH 7.9 with hydrochloric acid. The electrophoresis electrode solution was composed of 0.0892mol/litre Tris, 0.0890mol/litre boric acid and 2.5m mol/litre ETA. The electrophoresis was performed at a constant voltage of 200volt, while an upper electrode solution reservoir was filled with the electrode solution colored with BPB. Electric conduction was halted just when the dye front of BPB had reached the bottom of the gel. Staining was done using silver staining. After completion of electrophoresis, the gel was soaked for 20minutes in an aqueous solution containing 50%(v/v) methanol, 5%(w/v) trichloroacetic acid, and 3.5%(w/v) sulfosalicylic acid to fix the DNA, then rinsed twice with purified water for 15minutes. Thereafter the gel was soaked to be stained in a solution which was a mixture of two aqueous solutions in a proportion of 1:1, one solution contained 8%(w/v) sodium carbonate another solution contained 0.28%(v/v) formaldehyde, 1%(w/v) tungstosilicic acid, 0.02% (w/v) silver nitrate and 0.02%(w/v) ammonium nitrate. After a lapse of 7minutes during which the gel was stained to visualize the bands of separated molecules, the gel was removed from the dye solution, and immersed in 1%(v/v) acetic acid for 15minutes to stop the staining. The electrophoresis results proved the DNA bands clearly resolved. Table 7 below shows the mobility defined as the migration distance of a DNA fragment relative to the migration distance of BPB:

$$(\text{Mobility } \%) = (\text{Distance of each band from the lower end of well}) / (\text{Distance of the dye front of BPB from the lower end of well}) \cdot 100$$

[0066] Moreover, Table 8 is to explain the electrophoresis time for every gel sample.

Table 6

[Compositions of gel buffer solutions]			
	Sample-5	Sample-6	Sample-7
Tris (mol/litre)	0.094	0.082	0.080
Glycine (mol/litre)	0.192	0.167	0.167
Serine (mol/litre)	--	0.025	0.025
HCl (mol/litre)	0.080	0.0695	0.068
pH	7.5	titrated to 6.8 with acetic acid	titrated to 6.3 with acetic acid

Table 7

[Mobility of DNA fragments in gel just after produced]			
	Sample-5	Sample-6	Sample-7
Fragment size 495bp	25.91%	31.82%	31.78%
Fragment size 210bp	45.30%	53.79%	50.93%
Fragment size 162bp	53.79%	63.33%	59.35%
Fragment size 79bp	81.06%	89.39%	88.32%
Electrophoretic pattern	Clear	Clear	Clear

Table 8

[Electrophoresis time at constant voltage of 200V]		
Sample-5	Sample-6	Sample-7
49min.	52min.	50min.

[0067] As will be understood from Tables 6, 7 and 8, all the DNA fragments, when having undergone electrophoresis on the polyacrylamide gels with the gel solutions titrated to any pH of 6.3, 6.8 and 7.5, with using TBE, exhibited the electrophoretic patterns clearly resolved. With respect to the mobility and electrophoresis time, there was also no significant difference between any DNA fragments.

-Comparative Example 1-

[0068] With using the same glass plate cell as in Example 3 stated earlier, a polyacrylamide electrophoresis gel was, as with the gels in Example 3, prepared from monomer solution composed of 10% acrylamide(%T), 5% BIS(%C) and a buffer solution having a composition in concentration listed in Table 9. A commercially available marker DNA of known molecular weights was subjected to the electrophoresis using the polyacrylamide gel yielded as stated above. The electrophoresis results were satisfactory in migration distance and clearness of bands. The electrophoresis electrode solution used was the Ornstein and Davis electrophoresis electrode solution composed of 0.025mol/litre Tris and 0.192mol/litre glycine. In Table 10, the mobility is shown, which is defined as the ratio of the migration distance of the DNA fragment to the migration distance of the BPB, while Table 11 indicates the electrophoresis time.

Table 9

[Compositions of gel buffer solutions]	
	Comparative Sample-5
Tris (mol/litre)	0.080
Glycine (mol/litre)	0.172
Serine (mol/litre)	0.030
HCl (mol/litre)	0.068
pH	titrated to 6.3 with acetic acid

Table 10

[Mobility of DNA fragments in gel just after produced]	
	Comparative Sample-5
Fragment size 495bp	Unable to define
Fragment size 210bp	Unable to define
Fragment size 162bp	Unable to define
Fragment size 79bp	Unable to define
Electrophoretic pattern	Vague

Table 11 [Electrophoresis time at constant voltage of 20mA]

Comparative Sample-5
80min.

[0069] As will be seen in Tables 9, 10 and 11, all the DNA fragments, when having undergone electrophoresis on the polyacrylamide gels with the gel solutions titrated to pH of 6.3, with using the Ornstein and Davis electrophoresis electrode solution, exhibited no electrophoretic pattern clearly resolved, so that the individual DNA bands were unsatisfactorily diffuse.

-Example 4-

[0070] With using the same glass plate cell as in Example 3 stated earlier, a polyacrylamide electrophoresis gel was, as with the gels in Example 3, prepared from monomer solution composed of 10% acrylamide(%T), 5% BIS(%C) and a buffer solution having any one of composition in concentration listed below in Table 12.

[0071] The polyacrylamide gels prepared as stated above were stored at 5°C, and used every a preselected lapse of time for the runs in which the DNA fragments were subjected to electrophoresis using the same TBE as described in Example 3. The electrophoresis was run at a constant voltage of 200V per gel. The electrophoresis was allowed till the time 50minutes had gone by. The migration distance and the sharp pattern of bands were proved. The electrophoresis results are shown below in Tables 13, 14 and 15.

Table 12

[Compositions of gel buffer solutions]			
	Sample-8	Sample-9	Sample-10
Tris (mol/litre)	0.094	0.082	0.080
Glycine (mol/litre)	0.192	0.167	0.167
Serine (mol/litre)	--	0.025	0.025
HCl (mol/litre)	0.080	0.070	0.068
pH	7.5	titrated to 6.8 with acetic acid	titrated to 6.3 with acetic acid

Table 13

[Changes in mobility of DNA fragments in sample-8 after stored at 5°C for varying periods of time]				
	Soon after produced	After 2 months	After 4 months	After 6 months
Fragment size 495bp	30.80%	26.22%	17.72%	18.75%
Fragment size 210bp	52.85%	44.44%	35.04%	32.03%
Fragment size 162bp	62.36%	52.89%	42.91%	39.06%
Fragment size 79bp	90.87%	79.56%	70.47%	Unable to define
Electrophoretic pattern	Clear	Clear	Clear	Vague

Table 14

[Changes in mobility of DNA fragments in sample-9 after stored at 5°C for varying periods of time]				
	Soon after produced	After 2 months	After 4 months	After 6 months
Fragment size 495bp	26.20%	22.91%	23.25%	18.50%
Fragment size 210bp	47.16%	41.41%	41.67%	34.80%
Fragment size 162bp	55.90%	49.34%	50.00%	42.29%
Fragment size 79bp	85.59%	77.97%	78.51%	68.72%
Electrophoretic pattern	Clear	Clear	Clear	Clear

Table 15

[Changes in mobility of DNA fragments in sample-10 after stored at 5 °C for varying. periods of time]				
	Soon after produced	After 3 months	After 5 months	After 7 months
Fragment size 495bp	31.78%	31.05%	31.05%	30.52%
Fragment size 210bp	50.93%	49.61%	50.23%	50.23%
Fragment size 162bp	59.35%	57.99%	58.45%	59.15%
Fragment size 79bp	88.32%	87.21%	86.76%	87.79%
Electrophoretic pattern	Clear	Clear	Clear	Clear

[0072] As will be understood from Table 13, the polyacrylamide gel with the gel buffer solution of pH 7.5 was subject to degradation by hydrolysis with the lapse of time even at 5 °C, generating anionic groups, which behave in a manner that retards the mobility of DNA. After the lapse of six months, there was no detectable fragment of small molecule. Moreover, the Table 14 explains that the polyacrylamide gel with the gel buffer solution of pH 6.8 lowered much in the rate of hydrolysis compared with the gel buffer solution of pH 7.5. Nevertheless, much change in the mobility of DNA was seen in the gel after stored at 5 °C up to six months.

[0073] In contrast, the polyacrylamide gel with the gel buffer solution of pH 6.3, as shown in Table 15, experienced less changes in either of the mobility of DNA and the electrophoresis pattern even after having been stored at 5°C for seven months.

-Example 5-

[0074] A rectangular glass sheet of 12cm in width and 10cm long and another glass sheet of the same dimension made at an upper end thereof with a recess were assembled with a spacer of 1mm in thickness interposed between the confronting glass sheets to provide a glass plate cell. Monomer solutions were prepared, each of which was composed of a 10% acrylamide (%T) a 5% BIS (%C), and any one of buffer solutions composed as shown in Table 16 below. The monomer solutions were added and blended with 400ppm of APS and 400ppm of TEMED. The resultant mixture was poured into the glass plate cell where polymerization was allowed in a usual way to yield a polyacrylamide gel for electrophoresis.

[0075] A commercially available marker DNA of known molecular weights was subjected to electrophoresis using the polyacrylamide gel obtained as stated earlier. The electrophoresis results were satisfactory in migration distance and clearness of bands. The marker DNA, prior to use for electrophoresis test, was diluted one hundred times with a buffer solution composed of a 10m mol/litre Tris including 30%(w/v) sucrose and BPB of stainable level, 1m mol/litre ETA and 20m mol/litre sodium chloride solution, titrated to the pH 7.9 with hydrochloric acid. The electrophoresis electrode solution used was the Ornstein and Davis electrophoresis electrode solution composed of 0.025mol/litre Tris and 0.192mol/litre glycine. The electrophoresis was performed at a constant current of 20mA, while an upper electrode solution reservoir was filled with the electrode solution colored with BPB. Electric conduction was halted just when the dye front of BPB had reached the bottom of the gel. Staining was done using silver staining. After the completion of electrophoresis, the gel was soaked for 20minutes in an aqueous solution containing 50%(v/v) methanol, 5%(w/v) trichloroacetic acid, and 3.5%(w/v) sulfosalicylic acid to fix the DNA, then rinsed twice with purified water for 15minutes. Thereafter the gel was soaked to be stained in a solution, which was a mixture of two aqueous solutions in a proportion of 1:1, one solution contained 8%(w/v) sodium carbonate, another solution contained 0.28%(v/v) formaldehyde, 1% (w/v) tungstosilicic acid, 0.02%(w/v) silver nitrate and 0.02%(w/v) ammonium nitrate. After a lapse of 7minutes during which the gel was stained to visualize the bands of separated molecules, the gel was removed from the dye solution, and immersed in 1%(v/v) acetic acid for 15minutes to stop the staining. The electrophoresis results proved the DNA bands clearly resolved. Table 17 below shows the mobility defined as the migration distance of a DNA fragment relative to the migration distance of BPB:

$$(\text{Mobility } \%) = (\text{Distance of each band from the lower end of well}) / (\text{Distance of the dye front of BPB from the lower end of well}) \times 100$$

[0076] Moreover, Table 18 is to explain the electrophoresis time for every gel sample.

Table 16

[Compositions of gel buffer solutions]			
	Sample-11	Sample-12	Sample-13
Tris (mol/litre)	0.5	0.094	0.082
Glycine (mol/litre)	--	0.192	0.167
Serine (mol/litre)	--	--	0.025
HCl (mol/litre)	#	0.080	0.070
pH	# titrated to 8.8 with HCl	7.5	titrated to 6.8 with acetic acid

Table 17

[Mobility of DNA fragments in gel just after produced]			
	Gel Sample-11	Gel Sample-12	Gel Sample-13
Fragment size 495bp	33.03%	26.01%	26.03%
Fragment size 210bp	45.87%	45.98%	47.00%
Fragment size 162bp	52.29%	54.19%	55.46%
Fragment size 79bp	73.39%	81.55%	85.09%
Electrophoretic pattern	Clear	Clear	Clear

Table 18

[Electrophoresis time at constant current of 20mA]		
Gel Sample-11	Gel Sample-12	Gel Sample-13
74min.	80min.	79min.

[0077] As will be understood from Tables 16, 17 and 18, all the DNA fragments, when having undergone electrophoresis on the polyacrylamide gels with the gel solutions titrated to any pH of 7.5 and 6.8, with using the Ornstein and Davis electrophoresis electrode solution, exhibited the electrophoretic patterns clearly resolved. With respect to the mobility and electrophoresis time of the DNA fragments, there was also no significant difference, compared with the gel using the Ornstein and Davis electrophoresis electrode solution of pH 8.8.

[0078] In Tables 19 and 20 below, moreover, there are shown changes in the mobility in the samples-12 and 13 after having stored at 5°C.

Table 19

[Changes in mobility of DNA fragments in sample-12 after stored at 5 °C for varying periods of time]				
	Soon after produced	After 2 months	After 4 months	After 6 months
Fragment size 495bp	26.01%	22.48%	20.01%	14.69%
Fragment size 210bp	45.98%	38.35%	33.89%	26.48%
Fragment size 162bp	54.19%	45.45%	41.56%	34.25%
Fragment size 79bp	81.55%	67.12%	60.57%	Unable to define
Electrophoretic pattern	Clear	Clear	Clear	Vague

Table 20

[Changes in mobility of DNA fragments in sample-13 after stored at 5 °C for varying periods of time]				
	Soon after produced	After 2 months	After 4 months	After 6 months
Fragment size 495bp	26.03%	22.91%	20.84%	17.40%
Fragment size 210bp	47.00%	42.56%	40.50%	35.02%
Fragment size 162bp	55.46%	49.12%	47.56%	43.58%
Fragment size 79bp	85.09%	79.80%	76.12%	70.48%
Electrophoretic pattern	Clear	Clear	Clear	Clear

-Comparative Example 2-

[0079] With using the same glass plate cell as in Example 5, a polyacrylamide electrophoresis gel was, as with the gels in Example 5, prepared from monomer solution composed of 10% acrylamide(%T), 5% BIS(%C) and any one of buffer solutions having a composition in concentration listed in Table 21. A commercially available marker DNA of known molecular weights was subject to electrophoresis using the polyacrylamide gel yielded as stated above. The electrophoresis results were satisfactory in migration distance and clearness of bands. The electrophoresis electrode solution used was the Ornstein and Davis electrophoresis electrode solution composed of 0.025mol/litre Tris and 0.192mol/litre glycine. In Table 22, the mobility is shown, which is defined as the ratio of the migration distance of the DNA fragment to the migration distance of the BPB, while Table 23 indicates the electrophoresis time.

Table 21

[Compositions of gel buffer solutions]		
	Comparative Sample-6	Comparative Sample-7
Tris (mol/litre)	0.5	0.5
Glycine (mol/litre)	--	--
Serine (mol/litre)	--	--
HCl (mol/litre)	#	#
pH	titrated to 8.8 with HCl	titrated to 7.5 with HCl

Table 22

[Mobility of DNA fragments in gel just after having produced]		
	Comparative Sample-6	Comparative Sample-7
Fragment size 495bp	33.03%	49.32%
Fragment size 210bp	45.87%	74.43%
Fragment size 162bp	52.29%	85.39%
Fragment size 79bp	73.39%	--
Electrophoretic pattern	Clear	Clear

Table 23

[Electrophoresis time at constant current of 20mA]	
Comparative Sample-6	Comparative Sample-7
74min.	216min.

[0080] As will be understood from Tables 21, 22 and 23, when the gel buffer solution contained no glycine in the

solution, adjusted to pH 7.5 with Tris and hydrochloric acid, the electrophoresis gel rose in the potential gradient between the tris strong acid salt part and the tris weak acid salt part, so that the species undergoing separation were liable to focus on area neighboring the boundary. This made the fractionated molecular weight range reduced. Besides, the electrophoresis time was made elongated while analytic efficiency became worse.

[0081] Table 24 shows changes in mobility in the comparative gel sample-6 after storage at 5 °C for varying periods of time.

Table 24

[Changes in mobility of DNA fragments in comparative sample-6 after stored at 5°C for varying periods of time]				
	Soon after produced	After 2 months	After 4 months	After 6 months
Fragment size 495bp	33.03%	16.15%,	--	--
Fragment size 210bp	45.87%	21.58%	--	--
Fragment size 162bp	52.29%	32.41%	--	--
Fragment size 79bp	73.39%	Unable to define	--	--
Electrophoretic pattern	Clear	Vague	--	--

-Example 6-

[0082] With using the same glass plate cell as in Example 5 stated earlier, polyacrylamide electrophoresis gels were, as with the gels in Example 5, prepared from monomer solutions composed of 10% acrylamide(%T), 5% BIS(%C) and a buffer solution having any one composition of samples-12 and 13 in concentration listed earlier in Table 16. A commercially available marker DNA of known molecular weights was subjected to electrophoresis using the polyacrylamide gel obtained as stated earlier. The migration distance and the sharp pattern of bands were proved. The electrophoresis electrode solution used was the Laemmli's electrophoresis electrode solution containing 0.025mol/litre Tris, 0.192mol/litre glycine and 0.1%(w/v) SDS. The electrophoresis was run at a constant current of 20mA per gel. Table 25 shows the mobility representing a ratio of the migration distance of each DNA fragment to the migration distance of the BPB, while Table 26 explains the electrophoresis time per gel.

Table 25

[Mobility of DNA fragments in gel just after having produced]		
	Sample-12	Sample-13
Fragment size 495bp	25.99%	26.00%
Fragment size 210bp	45.93%	48.89%
Fragment size 162bp	54.10%	55.05%
Fragment size 79bp	81.00%	84.89%
Electrophoretic pattern	Clear	Clear

Table 26

[Electrophoresis time at constant current of 20mA]	
Sample-12	Sample-13
81min.	79min.

[0083] Tables 25 and 26 shown earlier prove that, even though the electrophoresis electrode solution used is the Laemmli's electrophoresis electrode solution containing SDS, which has been commonly used for the protein analysis, the gel prepared according to the present invention may serve well for producing sharp bands of the DNA fragments in analytic separation.

-Comparative Example 3-

[0084] With using the same glass plate cell as in Example 6, a polyacrylamide electrophoresis gel was, as with the gels in Example 6, prepared from a monomer solution composed of 10% acrylamide(%T), 5% BIS(%C) and a buffer solution having a composition in concentration described later in Table 27. A commercially available marker DNA of known molecular weights was subjected to electrophoresis using the polyacrylamide gel obtained as stated above. The migration distance and the sharp pattern of bands were proved. The electrophoresis electrode solution used was the Laemmli's electrophoresis electrode solution containing 0.025mol/litre Tris, 0.192mol/litre glycine and 0.1%(w/v) SDS. Table 28 shows the mobility representing a ratio of the migration distance of each DNA fragment to the migration distance of the BPB, while Table 29 explains the electrophoresis time.

Table 27

[Composition of gel buffer solution]	
	Comparative Sample-8
Tris (mol/litre)	0.5
Glycine (mol/litre)	--
Serine (mol/litre)	--
HCl (mol/litre)	#
pH	# titrated to 7.5 with HCl

Table 28

[Mobility of DNA fragments in gel just after having produced]	
	Comparative Sample-8
Fragment size 495bp	50.00%
Fragment size 210bp	75.43%
Fragment size 162bp	86.28%
Fragment size 79bp	--
Electrophoretic pattern	Clear

Table 29 [Electrophoresis time at constant current of 20mA]

Comparative Gel Sample-8
210min.

[0085] As shown in Tables 27, 28 and 29, when the DNA fragments were subjected to electrophoresis using the gel buffer solution containing no glycine in the solution, adjusted to pH 7.5 with Tris and hydrochloric acid, together with the Laemmli's electrophoresis electrode solution, the fractionated molecular weight range was rendered remarkably narrowed just like the run using electrophoresis using the Ornstein and Davis electrophoresis electrode solution. Moreover, the electrophoresis time was made elongated while analytic efficiency became worse.

Industrial Applicability

[0086] With the precast acrylamide electrophoresis gel in accordance with the present invention, the gel buffer solution contains both tris(hydroxymethyl)aminomethane and any ampholyte, titrated at a gel pH of from neutral to acidic range. This makes it possible to provide the electrophoresis gel that is retarded in hydrolysis to a significant extent,

thereby increased in useful shelf-life.

[0087] Thus, the precast acrylamide electrophoresis gel of the present invention can be stored for a prolonged period of time without loss of performance, so that it is possible to mass-produce in advance the precast gels differing diversely in separation characteristic or resolution from one another. The precast polyacrylamide gel of the present invention, when used in combination with the electrophoresis electrode solution applied extensively for the electrophoresis of proteins, serves well for the analytic separation of proteins and DNA and, when used in combination with the electrophoresis electrode solution commonly employed for nucleic acid, achieves the analytic separation of DNA.

[0088] Accordingly, the precast acrylamide electrophoresis gels of the present invention make it possible to save remarkably the labor and effort for analytic procedure, ensuring a large industrial applicability.

Claims

1. A precast polyacrylamide gel adapted for use in gel electrophoresis, in which an electrophoresis electrode solution is composed of an aqueous solution containing a tris(hydroxymethyl)aminomethane and an ampholyte, **characterized in that:**

(1) said tris(hydroxymethyl)aminomethane is present in a concentration of between 0.07mol/litre and 0.2mol/litre;

(2) said ampholyte is composed of glycine and at least one conjugate ampholyte,

(a) said conjugate ampholyte has a basic dissociation constant of $8.3 < pK_b < 9.6$,

(b) said conjugate ampholyte is added ranging in amount from 0.1 to 30mol% with respect to said glycine, and

(c) a total concentration of said conjugate ampholyte and said glycine ranges between 0.1 and 0.3mol/litre; and

(3) said electrophoresis electrode solution is adjusted to a pH of between pH 6.0 and pH 6.8.

2. A precast polyacrylamide electrophoresis gel recited in claim 1, **characterized in that** said conjugate ampholyte is an amino acid having the same number of anionic and cationic groups in a molecule.

3. A precast polyacrylamide electrophoresis gel recited in claim 1 or 2, **characterized in that** said conjugate ampholyte is any at least one selected from serine, glutamine, tryptophane, methionine and phenylalanine.

4. An use of a precast polyacrylamide electrophoresis gel **characterized in that** an analytic separation of proteins is performed using said precast gel recited in claims 1 to 3 in combination with said electrophoresis electrode solution containing tris(hydroxymethyl)aminomethane and glycine.

5. A process for fabricating a precast polyacrylamide electrophoresis, which is comprised of polymerizing an admixture of an acrylamide, polyfunctional crosslinking agent, water and a buffer solution of composition defined in the following (1) and (2) in the presence of a polymerization initiator under a condition of pH range 6.0 ~6.8:

(1) a tris(hydroxymethyl)aminomethane is present in a concentration of between 0.07mol/litre and 0.1mol/liter; and

(2) an ampholyte is composed of glycine and at least one conjugate ampholyte

(a) said conjugate ampholyte has a basic dissociation constant of $8.3 < pK_b < 9.6$,

(b) said conjugate ampholyte is added ranging in amount from 0.1 to 30mol% with respect to said glycine, and

(c) a total concentration of said conjugate ampholyte and said glycine ranges between 0.1 and 0.3mol/litre.

6. An use of a precast polyacrylamide electrophoresis gel wherein an analytic separation of proteins is performed using an electrophoresis electrode solution containing dodecyl sulfate, aside from tris(hydroxymethyl)aminomethane and glycine.

7. An electrophoresis system for DNA, which makes use of an electrophoresis electrode solution containing tris(hydroxymethyl)aminomethane and a precast polyacrylamide gel **characterized by** the followings (1) and (2);

(1) a gel buffer solution contained in said precast polyacrylamide gel contains the tris(hydroxymethyl)aminomethane, at least one ampholyte rendering glycine indispensable, and a monobasic acid, titrated to a pH of between pH 6.0 and pH 7.5, and

(2) said ampholyte contained in said gel buffer solution has a basic dissociation constant of $8.3 < pK_b < 9.6$, and includes an amino acid having the same number of anionic and cationic groups in a molecule.

8. An electrophoresis system for DNA recited in claim 7, wherein said gel buffer solution contains said monobasic acid consisting of at least any one of hydrochloric acid and acetic acid, titrated to a pH of between pH 6.0 and pH 6.5 to increase a useful shelf life of said precast gel.

9. An electrophoresis system for DNA recited in claim 7, wherein said gel buffer solution contains said monobasic acid consisting of at least any one of hydrochloric acid and acetic acid, titrated to a pH of between pH 6.5 and pH 7.5 to increase a useful shelf life of said precast gel.

10. An electrophoresis system for DNA recited in claim 7, wherein said electrophoresis electrode solution contains, aside from tris(hydroxymethyl)aminomethane, components defined in the following (1) and (2), titrated to a pH in a pH range 7.8 ~ 8.3:

(1) any one of acetic acid, phosphoric acid and boric acid; and

(2) a chelating agent of disodium ethylenediaminetetraacetate.

11. An electrophoresis system for DNA recited in claim 7 or 10, wherein said electrophoresis electrode solution containing said monobasic acid of boric acid.

12. An electrophoresis system for DNA recited in any one of claims 7, 10 and 11, wherein said electrophoresis electrode solution contains an ampholyte, aside from tris(hydroxymethyl)aminomethane, and titrated to a pH in a pH range 7.8~8.3.

13. An electrophoresis system for DNA recited in any one of claims 7, 10, 11 and 12, wherein said ampholyte contained in said electrophoresis electrode solution is glycine.

14. An electrophoresis system for DNA recited in any one of claims 7, 10, 11 and 12, wherein said electrophoresis electrode solution contains dodecyl sulfate, in addition to the ampholyte of glycine.

INTERNATIONAL SEARCH REPORT

International application No.

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A. CLASSIFICATION OF SUBJECT MATTER
Int. Cl.⁷ G01N27/447

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

Int. Cl.⁷ G01N27/447

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

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Kokai Jitsuyo Shinan Koho	1971-2000	Jitsuyo Shinan Toroku Koho	1996-2000

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

WPIL (DIALOG) : [IC=G01N27/447] *GLYCIN*TRIS?

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	WO, 96/16724, A (Pharmacia Biotech AB), 06 June, 1996 (06.06.96) & JP, 10-510363, A & EP, 794827, A & US, 6090252, A	1-14
A	WO, 97/04315, A (Electrophoretics International P.L.C.), 06 February, 1997 (06.02.97), & JP, 11-510044, A & EP, 842432, A & AU 6316496, A	1-14

☐ Further documents are listed in the continuation of Box C.☐ See patent family annex.

* Special categories of cited documents:

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- "&" document member of the same patent family

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